P-13

Insecticide susceptibility of the sand fly *Phlebotomus argentipes* in Bihar, India

<u>Ashish Shukla¹</u>, Rahul Chaubey^{1,2}, Anurag Kumar Kushwaha¹, Rajiv Kumar³, Om Prakash Singh⁴, Phillip Lawyer⁵, Christine Petersen^{6,7}, Scott Bernhardt⁸, Shyam Sundar^{1*}

1. Department of Medicine, Institute of Medical Sciences, Banaras Hindu University, India. 2. Kala Azar Medical Research Center, Muzaffarpur, India. 3. Centre of Experimental Medicine and Surgery, Institute of Medical Sciences, Banaras Hindu University, India. 4. Department of Biochemistry, Institute of Science, Banaras Hindu University, India. 5. Arthropod Collections, Monte L. Bean Life Science Museum, Brigham Young University, Provo, Utah, USA. 6. Department of Epidemiology, College of Public Health, University of Iowa. 7. Center for Emerging Infectious Diseases, University of Iowa, USA. 8. Department of Biology, Utah State University, Logan, Utah. *drshyamsundar@hotmail.com

Background

Leishmania donovani is a parasite that causes the parasitic disease visceral leishmaniasis in humans (HVL), which can be fatal if left untreated. HVL control in the Indian Subcontinent (ISC) has relied on the use of insecticides for Indoor Residual Spray (IRS). Alpha-cypermethrin, a synthetic pyrethroid, has taken the place of sand flies because they are now resistant to dichloro-diphenyl-trichloroethane (DDT) [1]. Alpha-cypermethrin is used to kill sand flies, thus increasing the risk of resistance development in sand flies under the pressure of regular exposure to insecticides [2,3]. Considering the possible development of resistance in the vector *Phlebotomus argentipes*, we monitored the susceptibility status of their field populations with exploration of probable underlying resistance mechanisms in villages with and without IRS.

Materials and methods

Sand flies were collected from 10 endemic villages in Muzaffarpur district, India. These villages were categorized as continuous IRS villages and discontinuous IRS villages based on their IRS histories. CDC light traps were installed in human dwellings, cattle sheds, and mixed dwellings. Captured sand flies were brought to the laboratory at Kala azar Medical Research Center (KAMRC), Muzaffarpur. *P. argentipes* were shorted and allowed for blood feeding and rearing as described previously [4]. This study was to determine the susceptibility status of field-caught sand flies and F1 progeny against the diagnostic doses (3µg/ml for 40 min) of alpha-cypermethrin using a CDC bottle bioassay. Polymorphisms in domain II segment 6 of the voltage-gated sodium ion channel (*VGSC*) gene of pyrethroid-supreme and tolerant *P. argentipes* were detected by DNA sequencing [5].

Result

We have observed 91.19% to 99.47% knockdown mortality and 89.34% to 98.93% mortality after 24 hours from field-caught sand flies and from F1 progeny, 91.7% to 98.89% and 90.16% to 98.33%, respectively. The lowest susceptibility levels were detected in vectors collected from the Sahebganj block, whereas the highest levels were detected in the Paroo block. Two non-synonymous mutations, L1014S and L1014F, were detected, which were found to be associated with pyrethroid/organochlorine resistance. The frequency of mutation is higher in villages having continuous IRS compared to discontinuous IRS villages.

Conclusion

Populations of *P. argentipes* in Bihar are largely susceptible to synthetic pyrethroids (alpha-cypermethrin). The susceptibility status of sand flies collected from the block showed that *P. argetipes* could potentially start developing resistance. The presence of insecticide target site insensitivity in a major proportion of flies implies potential future challenges for leishmaniasis control, suggesting the need for continuous monitoring and vigilance required to sustain the elimination once achieved.

Funding: supported by the Extramural Program of the National Institute of Allergy and Infectious Diseases, National Institute of Health (Tropical Medicine Research Center grant U19A1074321).

Conflict of interest:

The authors declare no conflict of interest.

References:

- Coleman M, Foster GM, Deb R, Pratap Singh R, Ismail HM, Shivam P, Ghosh AK, Dunkley S, Kumar V, Coleman M, Hemingway J. DDT-based indoor residual spraying suboptimal for visceral leishmaniasis elimination in India. Proceedings of the National Academy of Sciences. 2015 Jul 14;112(28):8573-8.b.
- 2. Pathirage DR, Karunaratne SP, Senanayake SC, Karunaweera ND. Insecticide susceptibility of the sand fly leishmaniasis vector Phlebotomus argentipes in Sri Lanka. Parasites & vectors. 2020 Dec;13:1-2.
- Roy L, Uranw S, Cloots K, Smekens T, Kiran U, Pyakurel UR, Das ML, S. Yadav R, Van Bortel W. Susceptibility status of the wild-caught Phlebotomus argentipes (Diptera: Psychodidae: Phlebotominae), the sand fly vector of visceral leishmaniasis, to different insecticides in Nepal. PLOS Neglected Tropical Diseases. 2022 Jul 14;16(7): e0010304.
- 4. Tiwary P, Singh SK, Kushwaha AK, Rowton E, Sacks D, Singh OP, Sundar S, Lawyer P. Establishing, expanding, and certifying a closed colony of Phlebotomus argentipes (Diptera: Psychodidae) for xenodiagnostic studies at the Kala Azar Medical Research Center, Muzaffarpur, Bihar, India. Journal of medical entomology. 2017 Sep 1;54(5):1129-39.
- Gomes B, Purkait B, Deb RM, Rama A, Singh RP, Foster GM, Coleman M, Kumar V, Paine M, Das P, Weetman D. Knockdown resistance mutations predict DDT resistance and pyrethroid tolerance in the visceral leishmaniasis vector *Phlebotomus argentipes*. PLoS neglected tropical diseases. 2017 Apr 17;11(4): e0005504.